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Proximity among protected area networks promotes functional connectivity for wintering waterfowl

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The equilibrium theorem provided a fundamental framework for understanding species' distributions and movement in fragmented ecosystems. Wetland-dependent avian species are model organisms to test insular predictions within protected area networks because their mobility allows surveillance of isolated patches without landscape barriers. We hypothesized size and isolation would influence functional connectivity of sanctuaries by GPS-marked wintering mallards (*Anas platyrhynchos*) within a mesocosm protected sanctuary area network. We evaluated functional connectivity and sanctuary use, measured by movements between sanctuaries, using a multistate modeling framework. Proximity drove connectivity, underscoring that patch isolation—not size—influenced connectivity, even for an avian species with no ascertainable landscape resistance or barriers. We also found that sanctuary use increased overwintering survival by reducing harvest mortality. Our test of equilibrium theory predictions demonstrated that isolation of protected sanctuary areas supersedes their size in determining functional connectivity for mallards and access to these areas may have direct fitness consequences. Our findings could refine land acquisition, restoration, and management practices with equal or greater emphasis on adjacency in protected area network design, especially for wetlanddependent migratory gamebirds.

Keywords *Anas platyrhynchos*, GPS telemetry, Functional connectivity, Island biogeography, Multistate modeling, National Wildlife Refuge System, Protected area networks, Sanctuary, Wintering waterfowl

The theory of island biogeography laid the foundation for critical insights across the fields of ecology, evolution, and conservation biology¹⁻³. In particular, the equilibrium model offered the first conceptual framework describing connectivity between discrete patches where bi-directional movement jointly depended on patch size and isolation^{4,5}. The model predicted increased colonization of larger and closer patches and emigration from smaller isolated patches^{6,7}. A key insight was its application to non-island environments where protected areas (i.e., "reserves", "refuges", "sanctuaries") become functional islands as land use change fragmented natural ecosystems⁸⁻¹¹. Conservation practitioners embraced these general principles for optimal design of protected area networks¹².

The influential single large versus several small (SLOSS) debate posited that a single large protected area promoted greatest species abundance, richness, and immigration than several smaller reserves^{13–17} (i.e., SL > SS). While global protected areas effectively conserve species diversity, abundance, and demography^{18–20}, the relative importance of size and connectivity are context-dependent. Conservation planners recognize influential factors of protected areas such as their authorized purposes (e.g., endangered species recovery vs. biodiversity goals), target organism(s) and traits (e.g., dispersal ability), and the surrounding landscape matrix^{2,21–24}. For instance, connectivity becomes more vital for recovery of endangered species or those with limited dispersal ability^{25,26}. Thus, protected area networks require case-specific evaluations based on the species and ecosystems they are designed to serve^{27–29}.

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Despite the contextual nature of protected area network design, large contiguous habitat is nearly always prioritized for protection over smaller areas^{30,31}, implying lower conservation value of small patches, which may undermine conservation or budgetary objectives^{32,33}. For example, megafauna require large protected areas but smaller connected patches may be as effective as large ones if they facilitate movement and dispersal (e.g., habitat corridors)^{22,34}. Likewise, conservation values of large marine protected areas are well-established (e.g., less sensitive to environmental perturbations)³⁵ but smaller marine protected area networks (i.e., stepping stones) may be equally effective depending on site characteristics, target taxonomic groups, and limiting consumptive use and other human disturbance^{36,37}. Smaller protected areas can also harbor substantial biodiversity, enhance landscape connectivity, are more cost-effective to acquire and maintain, and likely complement larger protected areas³⁸. Therefore, their potential conservation value should not be dismissed and may be especially important in the Anthropocene as natural ecosystems are increasingly fragmented and fiscal resources limited^{32,33,38}.

Wetland-dependent birds are notable models to evaluate protected area connectivity under the equilibrium framework because their mobility allows individuals to assess isolated patches without landscape barriers or resistance³⁹⁻⁴¹. Furthermore, wetland-dependent birds rely on severely threatened and fragmented ecosystems throughout their life-cycle⁴². Protected area wetlands provide resting and foraging areas during non-breeding seasons^{18,43}, migratory stopover and refueling sites⁴³⁻⁴⁷, and nesting and brood-rearing habitat^{48,49}. Ultimately, these sites are stepping stones that fulfil annual cycle requirements, and their connectivity is a prerequisite to the vitality and long-term viability of wetland-dependent migratory bird populations⁵⁰⁻⁵². Indeed, evaluations have emphasized a need for a greater number of integrated wetlands (i.e., complexes or networks) rather than larger contiguous wetland habitats to connect wetland-dependent bird movements at local and landscape scales and during different stages of their annual life cycle⁴³⁻⁴⁸.

Historically, global protected areas were established as sanctuaries (i.e., prohibited or very limited human access) for threatened or iconic species, landscapes, and seascapes to restore declining wildlife populations or promote biodiversity^{53,54}. Protected areas are increasing, but public support hinges on their utility to people and thus broader objectives are required to satisfy growing demands of increasingly diverse stakeholders^{55,56}. For example, the U.S. National Wildlife Refuge (NWR) System is the premiere example of a protected area network for wildlife conservation⁵⁷. In its infancy, the NWR System's mission was to protect land as inviolate sanctuary for at-risk and iconic wildlife⁵³ (e.g., brown pelican [Pelicanus occidentalis] and bald eagle [Haliaeetus leucocephalus]). However, NWRs now have unique designations (e.g., sanctuary, waterfowl production areas, human recreation) and subsequently, their authorized purposes change to meet public demand^{57,58}. One such directive for midcontinental NWR networks is to provide spatial sanctuary, free from hunting and other human disturbance, for migrating and wintering waterfowl with goals to: (1) provide rest areas and promulgate foraging resource requirements that promote population persistence; (2) serve as stepping stones that facilitate migratory and local wintering movements and connectivity; and (3) influence local-regional distributions of waterfow159-61. State agencies also establish smaller waterfowl sanctuaries to enhance disturbance-free wetland connectivity and waterfowl movement within sanctuary networks with implicit assumptions that protected sanctuary complexes enhance local waterfowl harvest opportunities and sustain abundant waterfowl populations regionally throughout autumn and winter⁶⁰⁻⁶².

Evaluating the effectiveness of protected area networks in meeting conservation objectives is challenging, especially for highly mobile species that occupy large geographic ranges and dynamic spatiotemporal distributions^{63,64}. Nevertheless, periodic and critical assessments are needed for effective management, restoration, or prioritization of new areas within or beyond established networks^{18,43,50,65}. Traditional evaluations of protected areas that span large spatial extents have recorded wildlife vital rates (e.g., abundance) or diversity indices^{18,66,67}, but they are historically limited by temporal frequency and therefore, an inability to directly measure connectivity among protected areas (*but see*⁶⁸). Emerging tracking technology allows practitioners to monitor movements among protected sanctuary areas directly, while removing spatial and temporal biases associated with resighting marked birds⁶⁸, thereby assessing functional connectivity and influential site characteristics (i.e., size, isolation) at biologically relevant spatial scales^{69–71}.

Our aim was to evaluate functional connectivity within a protected sanctuary network by wintering mallards (Anas platyrhynchos) by modeling daily movement transition probabilities to (i.e., immigration) and from (i.e., emigration) sanctuary "nodes"^{72,73}. Wintering mallards serve as an informative model species to test equilibrium predictions because their mobility allows aerial assessment of habitat patches with no landscape resistance⁷⁴. Yet, they rely on protected wetlands as suitable "islands" because of an otherwise inhospitable landscape matrix (i.e. intensive hunting). We hypothesized sanctuary size and isolation (i.e., distances) would influence movement transition probabilities among sanctuary nodes⁶. We predicted larger sanctuaries were local source populations and thus immigration transitions to larger sanctuaries were more likely, emigration transitions from larger sanctuaries were less likely, and the opposite immigration-emigration relationships for smaller sanctuaries. Likewise, we predicted mallards were more likely to transition to sanctuaries closer to one another compared to more distant sanctuaries. We also evaluated sanctuary use by mallards relative to capture-year and years after capture to ensure inferences were robust against transmitter marking biases (i.e., different sanctuary use behaviors in the first year compared to following winters). We predicted similar rates of sanctuary use between capture-year and return mallards and thus no or minimal marking biases. Last, we estimated overwintering survival for mallards that had access and used sanctuary compared to those that did not following capture. We hypothesized use of and access to protected sanctuary areas may confer fitness consequences; therefore, we predicted overwintering survival would be greater for mallards that used sanctuary because those that did not would experience greater harvest mortality. Our findings refine consequences of the equilibrium model, illustrating application and geographic generalizability for local, regional, and international sanctuary network design that promotes functional connectivity for a hunted gamebird during winter.

Results

We removed 3 mallards that migrated through but did not stay within our sanctuary network. We tracked 421 mallards (41% females, 24% juveniles) from 2019 to 2023, and 22 individuals had 2 or more winter seasons within the study region. Sixty-nine percent of mallards used 1 sanctuary node during winter, 19% used 2 nodes, and 12% used 3 or more (maximum = 8).

The probability of daily sanctuary transitions decreased as distance increased ($\beta = -0.11$; 90% CRI = -0.12to -0.10; Table 1). For every 10 km of spatial separation, mallards were 3.06 (90% CRI = 2.84-3.28; Fig. 1a) times less likely to transition from one sanctuary to another. Increased size of the "departure" (i.e., emigration) and "arrival" (i.e., immigration) sanctuaries increased the probability of sanctuary transitions (β s = 0.019 and 0.033; 90% CRIs = 0.009-0.032 and 0.021-0.045, respectively; Table 1); however, the magnitude of these effects were small (Fig. 1c,d). For example, given 10 km² (i.e., \bar{x} size) increase in emigration sanctuary, mallards were 1.22 (90% CRI = 1.09-1.38) times more likely to transition from one sanctuary to another (ψ range = 0.0002-0.006; Fig. 1c). Adults (β = 0.44; 90% CRI = 0.19-0.72) and males (β = 0.61; 90% CRI = 0.40 - 0.81) had greater transition probabilities than juveniles and females, respectively (Table 1). Adults were 1.56 (90% CRI = 1.21-2.05) times more likely to transition than juveniles, and males were 1.84 (90% CRI = 1.49-2.25) times more likely to transition than females (Fig. 1b). Probability of an adult male mallard making one or more transitions across the entire 120-day study period was 46.1% (90% CRI = 42.1-50.5%) when sanctuaries were 21.1 km apart (\bar{x} distance – 1 SD), 3.4% (90% CRI = 2.9-4.5%) when sanctuaries were 46.9 km apart (\bar{x}), and 0.2% (90% CRI = 0.1-0.3%) when sanctuaries were 72.7 km apart (\bar{x} + 1 SD).

Daily transition probabilities between sanctuary nodes was $\leq 6\%$ (Fig. 1; Table S1). The greatest probability of daily sanctuary transitions was between Reelfoot Lake NWR north to south and south to north units for adult males (ψ = 0.057 and 0.056; 90% CRIs = 0.049–0.065 and 0.048–0.064, respectively; Fig. 2). Adult male daily transitions were relatively high from Phillipy Refuge to Reelfoot Lake NWR north and south units ($\psi = 0.044$ and 0.036; 90% CRIs = 0.038-0.051 and 0.032-0.041, respectively) and Black Bayou Refuge to Reelfoot Lake NWR north and south units ($\psi = 0.033$ and 0.044; 90% CRIs = 0.029–0.037 and 0.039–0.050, respectively). In fact, most emigration-immigration combinations among sanctuaries with the greatest daily transition probabilities were within the "Reelfoot Wetlands Complex" because of the proximity of these sanctuaries to one another (Fig. 2). The "Upper Obion Wetland Complex", which included Bean Switch Refuge, Maness Swamp Refuge, and Hop-In Refuge, were also relatively well connected (Fig. 2). Among these, the greatest transition probabilities were from Hop-In Refuge to Maness Swamp Refuge ($\psi = 0.021$; 90% CRI = 0.019–0.024), from Maness to Hop-In (ψ = 0.020; 90% CRI = 0.018-0.023), from Bean Switch Refuge to Maness (ψ = 0.016; 90% CRI = 0.014-0.018) and from Maness to Bean Switch ($\psi = 0.015$; 90% CRI = 0.013–0.017). Although, farther away from the "Reelfoot Wetlands Complex", Lake Isom NWR was weakly connected with greatest connectivity from Lake Isom NWR to Reelfoot Lake NWR south unit ($\psi = 0.021$; 90% CRI = 0.019–0.023), to Black Bayou Refuge ($\psi = 0.012$; 90% CRI = 0.011–0.013), and to Reelfoot Lake NWR north (ψ = 0.010; 90% CRI = 0.008–0.013). In other words, Lake Isom NWR was an apparent source for the "Reelfoot Wetlands Complex". All other daily sanctuary transition probabilities were < 1% (Table S1; Fig. 2).

Mallards returning to the study region did not differ in number of sanctuaries used compared to those captured during winter ($\beta = 0.05$; SE = 0.28). Likewise, arrival or capture month did not affect sanctuary use by mallards (December: $\beta = -0.16$; SE = 0.003, January: $\beta = -0.33$; SE = 0.21, February: $\beta = -0.05$; SE = 0.34). However, the number of sanctuaries used increased with increasing time spent in the study region ($\beta = 0.015$; SE = 0.003). Specifically, mallards used 1.59 (90% CI 1.36–1.85) times as many sanctuaries for every 30 days in the region (Fig. 3).

Mallards that did not use sanctuary nodes following capture (11% or 45 individuals) had reduced overwintering survival compared to individuals that established winter ranges near and thus, had access to sanctuaries (Fig. S2). Specifically, individuals that used sanctuary had 3.06 (95% CI 1.77–5.31) times reduced hazard of death compared to individuals that never used sanctuary following capture. For 30 days within the sanctuary network, survival was 0.91 (95% CI 0.88–0.94) for mallards that had access and used sanctuary and 0.72 (95% CI 0.59–0.87) for mallards that did not use sanctuary. For 60 days, survival was 0.83 (95% CI 0.77–0.87) and 0.55

			90% credible intervals	
Hypothesis	Variable	β	Lower	Upper
System-specific	Intercept (sanctuary)	- 4.48	- 4.88	- 4.18
Individual characteristics	Age	0.44	0.19	0.72
	Sex	0.61	0.40	0.81
Island biogeography	Distance	- 0.11	- 0.12	- 0.10
	Size (emigration)	0.02	0.01	0.03
	Size (immigration)	0.03	0.02	0.05

Table 1. Parameter estimates on the logit scale (β) and associated 95% credible intervals from the multistate model evaluating daily sanctuary transition probabilities for system-specific (i.e., specific sanctuaries or the intercept), individual characteristic variables including age and sex, and island biogeography covariates including distance between sanctuaries and their sizes, including the departure or "emigration" sanctuary the individual left and the arrival or "immigration" sanctuary the individual transitioned to.

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Figure 1. The daily transition probabilities (ψ) from one waterfowl sanctuary to another by a wintering mallard (*Anas platyrhynchos*) captured or arriving in the west Tennessee and surrounding sanctuary complexes (November through February 2019–2023) relative to distance between sanctuaries (**a**), individual characteristics including female or male (green and orange, respectively) and age (juvenile or adult; **b**), and sanctuary sizes including the size of the sanctuary an individual left (emigration; **c**) and the size the individual transitioned to (immigration; **d**). Transition probabilities are associated with 68%, 90%, and 95% credible intervals for (**a**), (**c**), and (**d**) (dark to light gray) and 68% and 90% credible intervals for (**b**) (thick and skinny line, respectively). Predictions are generated from posterior distributions with all other values held constant at their mean value. Predictions for (**a**), (**c**), and (**d**) are for juvenile males because these were categorical indicator variables. Note different *y*-axes for visual aesthetics; despite increases or differences visually, distance between sanctuary nodes (**a**) was the only biologically meaningful effect. All Figures were produced in R version 4.3.3. https://www.r-project.org/.

(95% CI 0.39–0.77) for individuals that used and did not use sanctuary, respectively (Supplementary Context, Methods, and Results 2; Fig. S2).

Discussion

We evaluated functional connectivity of a highly mobile gamebird species within a mesocosm protected area network to refine applications of equilibrium theory. Proximity between sanctuary nodes promoted inter-patch movements more than area size, even for an avian species that is, theoretically, unimpeded by the matrix². Our findings align with previous research demonstrating isolation overrides patch size for connectivity outcomes, including for flying taxa^{27,75–78}. This highlights the equilibrium theory's assumption that landscape matrices impose dispersal costs, which may apply differentially to highly vagile or hunted species⁷⁹. For such mobile organisms, structural connectivity, facilitated by adjacent protected areas, can enhance functional connectivity without sole reliance on intervening habitat patches of lesser quality^{80,81}. Consequently, we suggest maximizing adjacency should be as much a focus as size for protected area networks aimed at increasing connectivity, especially for overwintering gamebirds and other wildlife that can transit above a hostile matrix to more suitable patches.



Figure 2. Predicted functional connectivity of mallards (Anas *platyrhynchos*) represented as daily sanctuary transition probabilities (*p*) among sanctuary nodes within the west Tennessee and surrounding sanctuary network of Arkansas, Kentucky, and Missouri. Individual mallards were captured and monitored with GPS transmitters from November through February 2019–2023. Sanctuary nodes included 4 National Wildlife Refuges: Big Lake National Wildlife Refuge (BLNWR) in Arkansas, Reelfoot Lake NWR north unit (RLNWR_N) in Kentucky and Tennessee, and Reelfoot Lake NWR south unit (RLNWR_S), Lake Isom NWR (LINWR), and Chickasaw NWRs in Tennessee. Additional smaller sanctuary nodes included state-owned waterfowl sanctuaries: Lake Lauderdale (LL), Horns Bluff (HB), White Lake (WL), Bean Switch (BS), Maness Swamp (M), Hop-in (HI), Black Bayou (BB), and Phillipy Waterfowl Refuges (P). Greatest functional connectivity was clearly within the Reelfoot Lake sanctuary nodes in the upper Obion River Complex including Hop-in, Bean Switch, and Maness Swamp Waterfowl Refuges also were more connected nodes illustrating distance, not size, as a primary driver of functional connectivity for wintering mallards. Figure was produced in R version 4.3.3. https://www.r-project.org/.

Measuring connectivity outcomes remains challenging and relies on indirect measures of genetic diversity, occupancy, or abundance⁸¹⁻⁸³. However, tracking individual movements provides a direct evaluation of network connectivity and possible barriers^{71,84,85}. Here, we used GPS tracks of mallards to estimate functional connectivity of protected sanctuary areas which revealed that individuals rarely transitioned between protected sanctuary nodes, despite the ability to fly above and avoid hunting risk when relocating (cf,⁸⁶), thereby implying some unknown costs. Critical to island biogeography is the assumption that the landscape matrix between suitable patches is inhospitable^{7,11}. While some wetlands beyond sanctuary borders may provide temporary refugia^{87,88}, our most connected area was also the most hunted (Table S1; Fig. S1). Instead, resource tracking and abundance theories predict reduced movement when resources are plentiful⁸⁹⁻⁹². Mallards likely foraged outside sanctuaries nocturnally when these patches were suitable and returned to sanctuary nodes diurnally⁹³⁻⁹⁶.

Few and proximity-biased transitions could be interpreted as energy conservation decision-making⁹⁷⁻⁹⁹. Indeed, waterfowl and other taxa minimize travel distances to foraging patches during winter unless payoffs at distant patches outweigh travel costs^{95,100-104}. However, food resources surrounding sanctuary nodes remained throughout winter precluding any need to conserve energy^{105,106}. Instead, few sanctuary transitions—predominantly to closer nodes—suggests adequate food resources within and around nodes, that translated into a single sanctuary being suitable the entire winter^{96,107}. A more likely cost of transitioning between sanctuaries is the immediate mortality risk by hunters^{18,60,108} (Fig. S2); that is, chronic hunting likely impeded connectivity. Mallards returning to the same sanctuary indicates a cognitive map of locally suitable patches¹⁰⁹⁻¹¹¹. Waterfowl in our region have only a short period to develop search images (i.e., pre-hunting season from arrival to ~ 5 December) and cognitive maps decay with time, in turn promoting shorter movement distances to areas frequently visited,



Figure 3. Predicted number of waterfowl sanctuary nodes used by wintering mallards (*Anas platyrhynchos*) within the west Tennessee and surrounding wetland complex protected sanctuary network relative to the number of days in the study area. Plots are faceted by the month (columns) and by individuals using sanctuaries during the same winter they were captured and individuals returning to the study area (rows). Figure was produced in R version 4.3.3. https://www.r-project.org/.

especially given diurnal movement constraints during hunting season^{96,100,109}. In other words, transitions to distant and unvisited sanctuaries would require exploratory behaviors that may increase hunter encounters^{108,112,113}. In concert, forage availability and abundance, spatial memory, and the negative fitness consequences for exploratory behavior (Supplementary Context, Methods, & Results) may explain why sanctuary proximity and not size promoted functional connectivity.

Authorized purposes for waterfowl spatial sanctuaries vary regionally, nationally, and internationally^{54,58}. Within our region, state-owned sanctuaries are intended to bolster or maintain local waterfowl abundance and facilitate movements among sanctuaries to improve waterfowl hunting and hunter satisfaction^{60,114}. National Wildlife Refuges in the region serve similar purposes but are six times the size of state-owned sanctuaries; therefore, they are better equipped to support biodiversity, population persistence, and host large abundances of waterfowl as local "source" populations to surrounding areas⁵³. However, our data indicate larger NWRs do not necessarily serve as local source populations that facilitate movement of mallards across our region, but we suggest they could if they were better connected to smaller state-owned sanctuary nodes within the network. Therefore, state conservation agencies that aim to increase waterfowl movements and connectivity should consider acquiring or leasing land that serves as stepping-stone sanctuaries to connect larger existing nodes, such as NWRs^{115,116}. A similar strategy was implemented in Louisiana, USA for northern pintails (A. acuta) with mixed results^{117,118}. Success or failure of attempts to improve wintering waterfowl connectivity undoubtedly depend on regional landscape matrices and sanctuary patch habitat quality³⁹. If food resources within and beyond sanctuary boundaries are abundant and hunting mortality risk in the surrounding matrix is high, waterfowl should minimize exploration to the extent physiologically possible, especially to distant nodes, making stepping-stone sanctuaries even more critical to improve functional connectivity^{102,107,115}. Additionally, smaller connecting sanctuaries must be disturbance-free^{87,119}. Hunting and other human disturbances within small sanctuaries would likely negate any positive connectivity benefits^{37,59,115}.

Private lands are crucial to wildlife conservation delivery worldwide¹²⁰⁻¹²² and can influence waterfowl resource selection and movement when protected legally^{18,43,123}. Protected private lands provide critical habitat and potential connectivity benefits among sanctuaries; however, single ownership parcels are often small and landowners typically recreate and disturb these areas, which likely limits their conservation value as stepping-stone sanctuaries. Private land cooperative partnerships may resolve this scalar problem as an effective mechanism for connecting waterfowl habitat while simultaneously improving recreational opportunities¹²⁴. Voluntary

partnerships among neighboring private landowners that collectively improve habitat quality and hunting experiences have proven effective at promoting connectivity for terrestrial wildlife species^{124–126}, but seldom has this model been translated to wetlands and waterfowl management. We suggest a similar model of private land conservation partnerships¹²⁷ to enhance waterfowl movement, landscape connectivity, and recreational opportunities. Private landowners may consider wetland management cooperatives (WMCs) that regulate waterfowl hunting temporally⁸⁸ and establish spatial sanctuary shared among the WMCs. This waterfowl management strategy has potential to enhance recreational opportunities by reducing isolation effects among state- and federally-owned sanctuary nodes thereby improving connectivity within the network.

Our findings refine applications of the equilibrium theory predictions for highly mobile and hunted species using protected area networks. Proximity promoted connectivity more than area, even for an unfettered avian migrant². Empirical studies like ours rarely support single large over several small reserves^{21,128,129}, yet conservation planners prioritize larger reserves, which may undermine landscape structural connectivity and disincentivize movement^{30,31}. For locally wintering waterfowl, numerous small sanctuaries could act as stepping stones to connect large reserves harboring source populations^{4,7,130}. In other words, "mainlands" likely already exist (e.g., National Wildlife Refuges) within established sanctuary networks for locally wintering waterfowl but functional connectivity may not. Conservation planners should consider the landscape matrix and species' movement transitions, distances, and range sizes when prioritizing areas for protection within pre-existing waterfowl or other protected area network designs^{43,69,70,131}. Publicly-funded programs to lease private lands as spatial sanctuaries or voluntary private land wetland management cooperatives (WMCs) that incorporate spatial sanctuary and limit hunting disturbance may enhance connectivity as stepping-stones within a sanctuary network. For example, if two 10 km² (3.9 mi²) sanctuaries were separated by 20 km (12.4 mi), a male mallard only had a 46% chance to transition to or from each sanctuary across the entire 120-day winter period. Hypothetically, should another 10 km² (3.9 mi²) stepping-stone sanctuary be established in the middle of the two existing sanctuaries (i.e., now 10 km or 6.2 mi between nodes), mallard functional connectivity would increase threefold, with a 88% probability of transitioning during the winter period.

Researchers should evaluate effectiveness (and nuances) of such programs aimed at increasing connectivity across waterfowl species and other wildlife. Simulations to reveal "optimal" connectivity thresholds are a logical extension to our work that would provide conservation planners with decision support for targeted land easements or acquisition¹³². Additionally, conservation agencies and their communication specialists may consider promoting potential benefits of private land cooperation to support wildlife connectivity in increasingly fragmented landscapes^{120,133,134}. Last, researchers should investigate a minimum sanctuary size to inform establishment of stepping-stone sanctuary sizes, which we could not identify because our smallest sanctuary node (1.3 km²) was well connected^{59,135}. Spatially-explicit models of hunting and waterfowl response to "disturbed areas" may be useful to infer risk perception and subsequently, inform minimum sanctuary sizes^{59,87,106} (Fig. S1).

Methods

Study system

Our study was conducted in west Tennessee and surrounding wetland complexes of west Kentucky, northeastern Arkansas, and southeastern Missouri, USA spanning 12,875 km² during autumn and winter 2019–2023. Waterfowl hunting is culturally and economically important to the region^{136–138}. Mallards are abundant and harvested intensively within and near the study region relative to the entire Mississippi Flyway of North America¹³⁹. Therefore, waterfowl sanctuaries provide needed spatially-defined and legally-designated safe and protected spaces for mallards and other waterfowl within an otherwise inhospitable landscape matrix (i.e., high hunter densities and activity across time and space^{60,96,114} (Fig. S1). Another purpose of the region's waterfowl sanctuaries is to maintain or enhance local–regional waterfowl abundance and facilitate movement among them during the waterfowl hunting season; both are assumed to increase harvest opportunity and hunter satisfaction^{60,140,141}.

Within this important geography exist four U.S. NWRs and seven state-owned waterfowl sanctuaries that vary in size and distance from one another. These sanctuaries prohibit hunting and other human activities on or before 15 November through 31 March^{60,96}. Intense hunting surrounding each waterfowl sanctuary in the region makes them functional "islands" among few other suitable habitats for waterfowl diurnally (Fig. S1)^{60,96}. Therefore, the region's sanctuary network is a model system or landscape mesocosm¹⁴² to test sanctuary network connectivity relative to sanctuary sizes and isolation (i.e., SL > SS) because it meets several criteria: (1) suitable patches (i.e., sanctuaries) are rare but with geographically¹⁴³ and biologically^{95,96} representative size-distance variation; (2) sanctuary patches are relatively homogenous spatially; and (3) the landscape matrix surrounding sanctuaries is "hostile" due to chronic anthropogenic hunting pressure (Fig. S1; Supplementary Context, Methods, Results 1)^{2,60,96}.

Animal capture and monitoring

We captured male and female mallards from October through February 2019–2022 on 9 of 11 waterfowl sanctuaries within our study region, thereby ensuring a spatially and temporally balanced sample. We banded ducks with U.S. Geological Survey aluminum tarsal bands and determined sex and age based on cloacal inversion, wing plumage, and bill color¹⁴⁴. We attached 20 g OrniTrack Global Positioning System-Global System for Mobile transmitters (GPS-GSM; Ornitela, UAB Švitrigailos, Vilnius, Lithuania) to birds weighing \geq 1000 g to ensure deployment packages remained below 3% of recommended body weights for unbiased monitoring¹⁴⁵. We programmed GPS-GSM transmitters to record hourly locations throughout the study. For analyses, we treated first year captured ducks and return wintering ducks as independent sampling units. All animal capture, handling procedures, and experimental protocols were in accordance with Tennessee Technological University's Institutional Animal Care and Use Committee protocol #19-20-002, authorized under Federal Banding Permit #05796, and adhered to ARRIVE guidelines (https://arriveguidelines.org).

Spatial and individual covariates

We used the protected area database (PAD-US; U.S. Geological Survey (USGS) Gap Analysis Project (GAP) 2022) to acquire U.S. NWR and state waterfowl sanctuary boundaries from northwest Tennessee, western Kentucky, eastern Arkansas, and Missouri. The PAD-US is an inventory of property boundaries with legal protected status intended to conserve biological diversity, recreation, and cultural uses. We defined spatial sanctuaries as areas managed for waterfowl and prohibited human recreation and disturbance before, during, and after the waterfowl hunting season. Following consultation with local biologists, we modified NWR and state-managed boundaries from PAD-US to exclude areas that allowed human recreation or other access, thereby ensuring our database only included waterfowl spatial sanctuaries⁴³. We also eliminated erroneous features from the analysis¹⁸ (e.g., boat docks, office buildings). Importantly, if sanctuaries were geographically separated—despite being considered one contiguous sanctuary—we separated them into two or more sanctuary nodes because mallards theoretically perceived these boundaries separately given the huntable landscape matrix between nodes (i.e., 1.6 and 2.1 km apart, respectively). Our resulting sanctuary network included five NWR nodes (three in Tennessee, one in Tennessee and Kentucky, and one in Arkansas) and eight state-owned waterfowl sanctuary nodes in Tennessee (n = 13).

For all federal and state-owned waterfowl sanctuaries, we calculated area (km²) and distance matrices (km) to and from each sanctuary using the *sf* package in R version $4.2.2^{146,147}$. Sanctuary area ranged from 1.3-45.7 km² ($\bar{x} = 9.7 \pm 11.8$ km²; n = 13) and minimum distances between sanctuaries ranged from 1.3-120.0 km ($\bar{x} = 46.9 \pm 25.8$; n = 78). We used sanctuary area and distances as covariates to test predictions that movement transitions (i.e., sanctuary departure and arrival) varied depending on the size of the emigrated sanctuary (e.g., source populations), the size of the sanctuary the individual relocated to, and the distance between them⁷. We also included age and sex of each individual as covariates to test predictions that males relocated more in search of limited females and pair bonding opportunities^{148,149} and juveniles relocated more because they were naïve to hunting risk implied by greater harvest rates^{150,151}.

Sanctuary transition multistate capture-recapture model

We developed multistate mark-recapture models in a Bayesian framework to estimate movement transition probabilities among sanctuaries^{152–154} (File S1, Table S1). We built daily encounter histories for each individual from 1 November through 29 February from 2019–2023 where we assigned categorical "states" (i.e., sanctuary node) for each mallard on each day during winter. We treated individuals with two or more winters as separate sampling units with separate encounter histories because individuals with > 1 year of data could begin their capture history at different sanctuary nodes (states) in winters $t + 1^{155}$. The encounter history for each individual began with their first GPS location on a sanctuary node, defined for each day between 1000 and 1900, which represented a time when most individuals would be within sanctuary boundaries^{94–96}. Additionally, we filtered the first 4 days of GPS locations following deployment to allow ducks to acclimate to GPS transmitters and harnesses⁹⁴. Individuals were assigned 1 of 14 possible states which included 13 sanctuary nodes and an unobserved state (File S1, Table S3, Table S4). Individuals with unobserved states missed GPS fixes or were not within sanctuary boundaries during the 1000–1900 observation window. Although not all sanctuary node transition combinations were reflected in the data, we did not restrict analyses to only observed transitions because all were biologically possible based on mallard movement and dispersal ability¹⁵⁵ (File S1, Table S3).

We estimated daily movement transition probabilities (ψ) from sanctuary node (*i*) to every other sanctuary (*j*), including the probability of staying on the current sanctuary, for a total of 169 transition probabilities ($\psi_{i,j}$), including "no transition". Specifically, we fitted generalized linear models with a multinomial link function to estimate movement probabilities on the logit scale for transitions from one sanctuary node (*s*) to any other node (*j*) for individual (*i*) relative to sanctuary sizes, distances, and age-sex demographics (1–13 possible transitions): $logit.\psi_{sji} = \beta_0 + \beta_{dist} \times Distance_{sj} + \beta_{size.s} \times Size_s + \beta_{size.j} \times Size_j + \beta_{sex} \times Sex_i + \beta_{age} \times Age_i$. We calculated the probability of staying on the current sanctuary node as $1 - \sum_{j=1}^{12} \psi_{s,j,i}$ ¹⁵⁴. In other words, the probability of transitioning to another sanctuary was estimated conditionally relative to distance between sanctuaries (km), sizes of the emigration and immigration sanctuaries (km²), and age and sex covariates. Conversely, the probability of staying (i.e., not transitioning) was calculated to ensure probabilities summed to 1¹⁵⁴. We estimated movement transition probabilities with one intercept (i.e., transitions from one sanctuary to any other sanctuary), as opposed to 13 sanctuary-specific intercepts^{154,156}.

We fitted the multistate model using Markov Chain Monte Carlo (MCMC) simulation with the *jagsUI* package in R^{147,157,158}. We specified vague priors for all model parameters¹⁵⁹. We used a parallel processing framework for computational efficiency which ran 10 independent Markov chains for 100 iterations each, a 20 iteration burn-in, and a thinning rate of 3¹⁶⁰. We proceeded with 100 iterations until models converged or until a priori maximum of 1000 iterations was reached. We monitored convergence based on visual inspection of the chains (Fig. 3) and the Gelman-Rubin statistic which converged $\hat{R} \le 1.01^{161}$. We reported coefficients on a logit scale and odds ratios with 90% credible intervals (CRI)¹⁶². We illustrated predicted relationships with 68%, 90%, and 95% CRI bands graphically¹⁶³.

Modeling sanctuary use and survival

We fitted a separate generalized linear model with a truncated Poisson error distribution and log-link function in a maximum likelihood framework using *glmmTMB* in R to estimate the effect of time spent in the study region (1–120 days), winter month (November, December, January, and February), and number of winters (1 winter = capture and > 1 winter = return) relative to the number of sanctuaries used^{147,164}. Intuitively, we predicted

the longer an individual remained within the study region, the greater number of sanctuaries used. We specified November as the categorical indicator and predicted individuals would use more sanctuaries during November and February and fewer sanctuaries during December and January to minimize risk of hunting mortality (i.e., less movement during hunting season). Importantly, we included the number of winters an individual was monitored (i.e., 1 or > 1) to ascertain possible confounding sanctuary use behavior due to marker effects. Specifically, we assumed that an individual that returned to the region and thus survived the previous winter–fall migration was reasonably unaffected by their transmitter; therefore, no clear statistical difference of sanctuary use between capture-year wintering ducks compared to ducks that returned to the study region suggests no behavioral biases of sanctuary use as a result of external transmitters. We calculated Pearson's correlation (r) between pairs of continuous covariates but none were correlated ($|r| \le 0.6$)¹⁶⁵. We used the *DHARMa* package in R to ensure model assumptions were not violoted¹⁶⁶ (Fig. S4). We provided estimated coefficients and considered covariates statistically significant if 90% confidence intervals did not bound zero^{167,168}.

We identified 45 individuals (11%) that never used sanctuary following capture (Supplementary Context, Methods, Results 2). We assigned a binary independent variable of sanctuary use to each individual where "0" denoted individuals that never used a sanctuary node following capture and "1" was the opposite. We fitted a Cox proportional hazard model in the *survival* package in R to estimate known-fate Kaplan-Meir survival curves and hazard ratios^{169,170}. We right-censored individuals that did not die by the end of winter (120-days from November–February) or stopped transmitting, assuming unbiased censoring^{171,172}. We tested whether overwintering survival was different between sanctuary and non-sanctuary mallards using a log-rank χ^2 test at an a priori $\alpha = 0.05^{173}$. We reported a seasonal baseline hazard ratio, survival probabilities at 30 and 60 days and associated variance estimates for each GPS-marked cohort. Additional details are provided in Supplementary Context, Methods, Results 2.

Animal ethics

Duck capture and handling procedures were in accordance with Tennessee Technological University's Institutional Animal Care and Use Committee protocol #19-20-002 and authorized under Federal Banding Permit #05796.

Data availability

Relevant data and code are included in Supporting Information and are available in GitHub and published through Zenodo at https://doi.org/10.5281/zenodo.10150699.

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References

- 1. Losos, J. B. & Ricklefs, R. E. The Theory of Island Biogeography Re-visited (Princeton University Press, 2010).
- 2. Fahrig, L. et al. Resolving the SLOSS dilemma for biodiversity conservation: A research agenda. Biol. Rev. 97, 99–114 (2022).
- Whittaker, R. J. & Fernández-Palacios, J. M. Island Biogeography: Ecology, Evolution, and Conservation 2nd edn. (Oxford University Press, 2007).
- Levins, R. Some demographic and genetic consequences of environmental heterogeneity for biological control. Bull. Entomol. Soc. Am. 15, 237–240 (1969).
- 5. MacArthur, R. H. & Wilson, E. O. An equilibrium theory of insular zoogeography. Evolution 17, 373-387 (1963).
- 6. Levins, R. Extinction. Lectures in Mathematics and Life Sciences 2, 77–107 (1970).
- 7. MacArthur, R. H. & Wilson, E. O. The Theory of Island Biogeography (Princeton University Press, 1967).
- 8. Diamond, J. M. The island dilemma: Lessons of modern geographic studies for the design of natural reserves. *Biol. Conserv.* 7, 129–146 (1975).
- Haddad, N. M. et al. Habitat fragmentation and its lasting impact on Earth's ecosystems. Sci. Adv. https://doi.org/10.1126/sciadv. 1500052 (2015).
- Laurance, W. F. Beyond island biogeography theory: Understanding habitat fragmentation in the real world. In *The Theory of Island Biogeography Revisited*, 214–236. http://www.jstor.org/stable/j.ctt7s5m6.13 (Princeton University Press, 2010).
- Turner, M. G., Gardner, R. H. & Oniell, R. V. Landscape Ecology in Theory and Practice 406 (Springer, 2001). https://doi.org/10. 1007/b97434.
- 12. Trianstis, K. A. & Bhagwat, S. A. Applied island biogeography. In *Conservation Biogeography* (eds Ladle, R. J. & Whittaker, R. J.) (Blackwell Publishing Ltd, 2011).
- 13. Diamond, J. M. Island biogeography and conservation: Strategy and limitations. Science 193, 1027–1029 (1976).
- Quinn, J. F. & Harrison, S. P. Effect of habitat fragmentation and isolation on species richness: Evidence from biogeographic patterns. *Oecologia* 75, 132–140 (1988).
- 15. May, R. M. Island biogeography and the design of wildlife preserves. *Nature* 254, 177–178 (1975).
- 16. Simberloff, D. S. & Abele, L. G. Island biogeography theory and conservation practice. *Science* **191**, 285–286 (1976).
 - Simberloff, D. S. & Abele, L. G. Refuge design and Island biogeographic theory: Effects of fragmentation. Am. Nat. 120, 41–50 (1982).
 - Masto, N. M. *et al.* Waterbird-habitat relationships in South Carolina: Implications for protection, restoration, and management of coastal and inland wetlands. *Restor. Ecol.* 31, e13956. https://doi.org/10.1111/rec.13956 (2023).
- Rodrigues, A. S. L. *et al.* Effectiveness of global protected area network in representing species diversity. *Nature* 428, 640–643 (2004).
- Soriando-Redondo, A. *et al.* Demographic rates reveal the benefits of protected areas in a long-lived migratory bird. *Proc. Natl.* Acad. Sci. 120, e2212035120 (2023).
- 21. Fahrig, L. Why do several small patches hold more species than few large patches?. Glob. Ecol. Biogeogr. 29, 615-628 (2020).
- 22. Margules, C. R. & Pressey, R. L. Systematic conservation planning. *Nature* 405, 243–253 (2000).
- 23. Ricketts, T. H. The matrix matters: Effective isolation in fragmented landscapes. Am. Nat. 158, 87-99 (2001).
- 24. Sarkar, S. Complementarity and the selection of nature reserves: Algorithms and the origins of conservation planning, 1980–1995. *Arch. Hist. Exact Sci.* **66**, 397–426 (2012).
- 25. Banks, S. C. et al. How does ecological disturbance influence genetic diversity?. Trends Ecol. Evol. 28, 670-679 (2013).

- 26. Doerr, V. A. J. et al. Designing landscapes for biodiversity under climate change. Ecol. Appl. 21, 1989–1996 (2011).
- 27. Fahrig, L. et al. Is habitat fragmentation bad for biodiversity?. Biol. Conserv. 230, 179-186 (2019).
- Jantke, K. et al. Is habitat connectivity relevant for protected areas? A case study of plants, mammals and birds. Divers. Distrib. 24, 730-742 (2018).
- Saura, S. et al. Protected area connectivity: Shortfalls in global targets and country-level priorities. Biol. Conserv. 209, 349–357 (2017).
- Armsworth, P. R. *et al.* Is conservation right to go big? Protected area size and conservation return-on-investment. *Biol. Conserv.* 225, 229–236 (2018).
- 31. Hill, M. et al. New policy directions for global pond conservation. Conserv. Lett. 11, e12447 (2018).
- Ribeiro, M. C., Metzger, J. P., Martensen, A. C., Ponzoni, F. J. & Hirota, M. M. The Brazilian Atlantic Forest: How much is left, and how is the remaining forest distributed? Implications for conservation. *Biol. Conserv.* 142, 1141–1153 (2009).
- Walker, J. J. *et al.* Distribution of duck broods relative to habitat characteristics in the Prairie Pothole Region. *J. Wildl. Manag.* 77, 392–404. https://doi.org/10.1002/jwmg.466 (2013).
- Sage, A. H., Hillis, V., Graves, R. A., Burnham, M. & Carter, N. H. Paths of coexistence: Spatially predicting acceptance of grizzly bears along key movement corridors. *Biol. Conserv.* 266, 109–468. https://doi.org/10.1016/j.biocon.2022.109468 (2022).
- 35. Lester, S. E. et al. Biological effects within no-take marine reserves: A global synthesis. Mar. Ecol. Prog. Ser. 384, 33-46 (2009).
- Lee, K. A., Huveneers, C., Macdonald, T. & Harcourt, R. G. Size isn't everything: Movements, home range, and habitat preferences of eastern blue gropers (*Achoerodus viridis*) demonstrate the efficacy of a small marine reserve. *Aquat. Conserv. Mar. Freshw. Ecosyst.* 25(2), 174–186 (2015).
- Turnbull, J. W. et al. Key drivers of effectiveness in small marine protected areas. Biodivers. Conserv. 27, 2217–2247. https://doi. org/10.1007/s10531 (2018).
- Volenec, Z. M. & Dobson, A. P. Conservation values of small reserves. Conserv. Biol. 34, 66–79. https://doi.org/10.1111/cobi. 13308 (2020).
- Baguette, M., Blanchet, S., Legrand, D., Stevens, V. M. & Turlure, C. Individual dispersal, landscape connectivity and ecological networks. *Biol. Rev.* 88(2), 310–326 (2013).
- Goncalves, J., Honrado, J. P., Vicente, J. R. & Covantos, E. A model-based framework for assessing the vulnerability of low dispersal vertebrates to landscape fragmentation under environmental change. *Ecol. Complex.* 28, 174–184. https://doi.org/10. 1016/j.ecocom.2016.05.033 (2016).
- Warren, M. J., Wallin, D. O., Beausoleil, R. A. & Warheit, K. I. Forest cover mediates genetic connectivity of northwestern cougars. Conserv. Genet. 17, 1011–11024. https://doi.org/10.1007/s10592-016-0840-7 (2016).
- Davidson, N. C. How much wetland has the world lost? Long-term and recent trends in global wetland area. *Mar. Freshw. Res.* 65(10), 934–941. https://doi.org/10.1071/MF14173 (2014).
- Beatty, W. S. et al. The role of protected area wetlands in waterfowl habitat conservation: Implications for protected area network design. Biol. Conserv. 176, 144–152 (2014).
- 44. Beatty, W. S. *et al.* Landscape effects on mallard habitat selection at multiple spatial scales during the non-breeding period. *Landsc. Ecol.* **29**, 989–1000 (2014).
- McCulloch, G., Aebischer, A. & Irvine, K. Satellite tracking of flamingos in southern Africa: The importance of small wetlands for management and conservation. Oryx 37, 480–483. https://doi.org/10.1017/S003060530300851 (2003).
- 46. Myers, J. P. et al. Conservation strategy for migratory species. Am. Sci. 75, 19–26 (1987).
- Sanders, F. J., Handmaker, M. C., Johnson, A. S. & Senner, N. R. Nocturnal roost on South Carolina coast supports nearly half of Atlantic coast population of Hudsonian Whimbrel *Numenius hudsonicus* during northward migration. *Wader Study* https:// doi.org/10.18194/ws.00228 (2023).
- Stephens, S. E., Rotella, J. J., Lindberg, M. S., Taper, M. L. & Ringelman, J. K. Duck nest survival in the Missouri Coteau of North Dakota: Landscape effects at multiple spatial scales. *Ecol. Appl.* 15, 2137–2149 (2005).
- Mitchell, B. J. et al. Wetland occupancy by duck broods in cropland-dominated landscapes of the United States Prairie Pothole Region. J. Wildl. Manag. 87, e22347. https://doi.org/10.1002/jwmg.22347 (2023).
- 50. Xu, Y. *et al.* Loss of functional connectivity in migration networks induces population decline in migratory birds. *Ecol. Appl.* **29**, e01960. https://doi.org/10.1002/eap.1960 (2019).
- Xu, Y., Kieboom, M., van Lammeren, R. J. A., Si, Y. & de Boer, W. F. Indicators of site loss from a migration network: Anthropogenic factors influence waterfowl movement patterns at stopover sites. *Glob. Ecol. Conserv.* 25, e01435. https://doi.org/10.1016/j.gecco.2020.e01435 (2021).
- 52. Xu, Y. *et al.* Beyond site-specific criteria: Conservation of migratory birds and their habitats from a network perspective. *Diversity* 14, 353. https://doi.org/10.3390/d14050353 (2022).
- Davison, R. P., Falcucci, A., Maiorano, L. & Scott, J. M. The National Wildlife Refuge System. *Endangered Species Act* 30(1), 90–100 (2006).
- Watson, J. E., Dudley, N., Segan, D. B. & Hockings, M. The performance and potential of protected areas. *Nature* 515, 67–73. https://doi.org/10.1038/nature13947 (2014).
- Naughton-Treves, L., Holland, M. B. & Brandon, K. The role of protected areas in conserving biodiversity and sustaining local livelihoods. *Annu. Rev. Environ. Resour.* 30, 219–252 (2005).
- 56. Griffith, B. et al. Climate change adaptation for the US National Wildlife Refuge System. Environ. Manag. 44, 1043-1052 (2009).
- 57. Fischman, R. L. The National Wildlife Refuge System and the hallmarks of modern organic legislation. *Ecol. Law Q.* 29, 457–622 (2002).
- Fischman, R. L. From words to action: The impact and legal status of the 2006 National Wildlife Refuge System management policies. Stanf. Environ. Law J. 25, 77–135 (2007).
- Fox, A. D. & Madsen, J. Behavioral and distributional effects of hunting disturbance on waterbirds in Europe: Implications for refuge design. J. Appl. Ecol. 34, 1–13 (1997).
- Blake-Bradshaw, A. G. *et al.* Influence of sanctuary disturbance, weather, and landscape characteristics on waterfowl harvest opportunity in western Tennessee. J. Wildl. Manag. 2, 24–70. https://doi.org/10.1002/jwmg.22470 (2023).
- 61. Evans, D. M. & Day, K. R. Hunting disturbance on a large shallow lake: The effectiveness of waterfowl refuges. *Ibis* **144**, 2–8 (2002).
- Guillemain, M., Martin, G. R. & Fritz, H. Feeding methods, visual fields, and vigilance in dabbling ducks (*Anatidae*). *Funct. Ecol.* 16, 522–529 (2002).
- 63. Leverington, F., Costa, K. L., Pavese, H., Lisle, A. & Hockings, M. A global analysis of protected area management effectiveness. *Environ. Manag.* **46**, 685–698 (2010).
- Runge, C. A., Martin, T. G., Possingham, H. P., Willis, S. G. & Fuller, R. A. Conserving mobile species. Front. Ecol. Environ. 12, 395–402 (2014).
- Leonard, P. B. et al. Landscape connectivity losses due to sea level rise and land use change. Anim. Conserv. https://doi.org/10. 1111/acv.12289 (2016).
- 66. Gaget, E. *et al.* Benefits of protected areas for nonbreeding waterbirds adjusting their distributions under climate warming. *Conserv. Biol.* **35**, 834–845 (2021).

- 67. Pearse, A. T., Kaminski, R. M., Reinecke, K. J. & Dinsmore, S. J. Local and landscape associations between wintering dabbling ducks and wetland complexes in Mississippi. *Wetlands* **32**, 859–869 (2012).
- Ferraz, G. et al. A large-scale deforestation experiment: Effects of patch area and isolation on amazon birds. Science 315, 238–241. https://doi.org/10.1126/science.ll33097 (2007).
- Allen, A. M. & Singh, N. J. Linking movement ecology with wildlife management and conservation. Front. Ecol. Evol. 3, 155. https://doi.org/10.3389/fevo.2015.00155 (2016).
- Franco, A. D. *et al.* Linking home ranges to protected area size: The case study of the Mediterranean Sea. *Biol. Conserv.* 221, 175–181. https://doi.org/10.1016/j.biocon.2018.03.012 (2018).
- Stewart, F. E., Darlington, S., Volpe, J. P., McAdie, M. & Fisher, J. T. Corridors best facilitate functional connectivity across a protected area network. *Sci. Rep.* 9, 10852. https://doi.org/10.1038/s41598-019-47067-x (2019).
- Taylor, P. D., Fahrig, L., Henein, K. & Merriam, G. Connectivity is a vital element of landscape structure. *Oikos* 1, 571–573 (1993).
 Fahrig, L., Arroyo-Rodríguez, V., Cazetta, E., Ford, A., Lancaster, J., & Ranius, T. Landscape connectivity. In *Routledge Handbook of Landscape Ecology*, 67–88, https://doi.org/10.4324/9780429399480-5 (2021).
- Martin, G. R. Through birds' eyes: Insights into avian sensory ecology. J. Ornithol. 153, 23–48. https://doi.org/10.1007/s10336-011-0771-5 (2012).
- Concepción, E. D., Díaz, M. & Baquero, R. Effects of landscape complexity on the ecological effectiveness of agri-environment schemes. Landsc. Ecol. 23, 135–148 (2008).
- Martensen, A. C., Pimentel, R. G. & Ribeiro, M. C. Relative effects of fragment size and connectivity on bird community in the Atlantic Rain Forest: Implications for conservation. *Biol. Conserv.* 141, 2184–2192 (2008).
- 77. Moilanen, A. & Hanski, I. Metapopulation dynamics: Effects of habitat quality and landscape structure. *Ecology* **79**, 2503–2515 (1998).
- 78. Newmark, W. D. Isolation of African protected areas. Front. Ecol. Environ. 6, 321-328 (2008).
- 79. Baguette, M. & Van Dyck, H. Landscape connectivity and animal behavior: Functional grain as a key determinant for dispersal. *Landsc. Ecol.* **22**, 1117–1129 (2007).
- Cushman, S., McRae, A., Adriaensen, B., Beier, F., Shirley, P., & Zeller, M. Biological corridors and connectivity. In *Key Topics in Conservation Biology*, 384–403 (2013).
- Zeller, K. A., McGarigal, K. & Whiteley, A. R. Estimating landscape resistance to movement: A review. Landsc. Ecol. 27, 777–797 (2012).
- LaPoint, S., Gallery, P., Wikelski, M. & Kays, R. Animal behavior, cost-based corridor models, and real corridors. *Landsc. Ecol.* 28, 1615–1630 (2013).
- Rudnick, D. A. *et al.* The role of landscape connectivity in planning and implementing conservation and restoration priorities. *Issues Ecol.* 16, 1–20 (2012).
- Cushman, S. A. & Lewis, J. S. Movement behavior explains genetic differentiation in American black bears. Landsc. Ecol. 25, 1613–1625 (2010).
- McClure, C. J., Rolek, B. W. & Hill, G. E. Predicting occupancy of wintering migratory birds: Is microhabitat information necessary?. Condor 114, 482–490 (2012).
- Rio-Maior, H., Nakamura, M., Álvares, F. & Beja, P. Designing the landscape of coexistence: Integrating risk avoidance, habitat selection and functional connectivity to inform large carnivore conservation. *Biol. Conserv.* 235, 178–188. https://doi.org/10. 1016/j.biocon.2019.04.021 (2019).
- Hagy, H. M., Horath, M. M., Yetter, A. P., Hine, C. S. & Smith, R. V. Evaluating tradeoffs between sanctuary for migrating waterbirds and recreational opportunities in a restored wetland complex. *Hydrobiologia* https://doi.org/10.1007/s10750-016-2711-0 (2017).
- St. James, E. A., Schummer, M. L., Kaminski, R. M., Burger, L. W. & Penny, E. J. Effect of weekly hunting frequency on duck abundances in Mississippi Wildlife Management areas. J. Southeastern Assoc. Fish Wildl. Agencies 4, 144–150 (2013).
- Fretwell, D. S. & Lucas, H. L. J. On territorial behavior and other factors influencing habitat distribution in birds. Acta Biotheoretica 19, 16–36 (1969).
- 90. Ford, R. G. Home range in a patchy environment: Optimal foraging predictions. Am. Zool. 23(2), 315-326 (1983).
- Johnson, D. H. The comparison of usage and availability measurements for evaluating resource preference. *Ecology* 61, 65–71. https://doi.org/10.2307/1937156 (1980).
- 92. Morris, D. W. How can we apply theories of habitat selection to wildlife conservation and management?. *Wildl. Res.* **30**, 303–319 (2003).
- Gaynor, K. M., Hojnowski, C. E., Carter, N. H. & Brashares, J. S. The influence of human disturbance on wildlife nocturnality. Science 360, 1232–1235 (2018).
- Palumbo, M. D., Petrie, S. A., Schummer, M., Rubin, B. D. & Bonner, S. Mallard resource selection trade-offs in a heterogeneous environment during autumn and winter. *Ecol. Evol.* 9, 1798–1808. https://doi.org/10.1002/ece3.4864 (2019).
- Shirkey, B. T., Palumbo, M. D. & Simpson, J. W. Land cover switching in autumn by female mallards in Ohio. J. Wildl. Manag. 84, 968–978 (2020).
- 96. Masto, N. M. Human access constrains optimal foraging and habitat availability in an avian generalist. *Ecol. Appl.* (in press) (2024).
- Gallagher, A. J., Creel, S., Wilson, R. P. & Cooke, S. J. Energy landscapes and the landscape of fear. *Trends Ecol. Evol.* 32, 88–96. https://doi.org/10.1016/j.tree.2016.10.010 (2017).
- 98. Heath, J. P. & Gilchrist, H. G. When foraging becomes unprofitable: Energetic of diving in tidal currents by common eiders wintering in the Arctic. *Mar. Ecol. Prog. Ser.* **403**, 279–290 (2010).
- Mosser, A. A., Avgar, T., Brown, G. S., Walker, S. & Fryxell, J. M. Towards an energetic landscape: Broad-scale accelerometry in woodland caribou. J. Anim. Ecol. 83, 916–922 (2014).
- Chudzinska, M. et al. AgentSeal: Agent-based model describing movement of marine central-place foragers. Ecol. Model. 440, 109–397 (2021).
- Lamb, J. S., Satge, Y. G. & Jodice, P. G. R. Seasonal variation in environmental and behavioural drivers of annual-cycle habitat selection in a nearshore seabird. *Divers. Distrib.* 26, 254–266. https://doi.org/10.1111/ddi.13015 (2020).
- Legagneux, P., Blaize, C., Latraube, F., Gautier, J. & Bretagnolle, V. Variation in home-range size and movements of wintering dabbling ducks. J. Ornithol. 150, 183–193 (2009).
- 103. McDuie, F. *et al.* GPS tracking data reveals daily spatio-temporal movement patterns of waterfowl. *Mov. Ecol.* **7**, 6. https://doi.org/10.1185/s40462-019-0146-8 (2019).
- McDuie, F. et al. Informing wetland management with waterfowl movement and sanctuary use responses to human-induced disturbance. J. Environ. Manag. 297, 113–170 (2021).
- Highway, C. J. Spatiotemporal factors influencing mallard foraging and activity dynamics. M.S. Thesis, Tennessee Technological University, Cookeville, TN, USA (2022).
- Masto, N. M. Landscape and behavioral ecology of mallards during the nonbreeding season. Ph.D. Dissertation, Tennessee Technological University, Cookeville, TN, USA (2023).
- Guillemain, M., Mondain-Monval, J. Y., Weissenbacher, E., Brochet, A. L. & Olivier, A. Hunting bag and distance from nearest day-roost in Camargue ducks. Wildl. Biol. 14, 379–385 (2008).

- Palumbo, M. D., Petrie, S. A., Schummer, M. L., Rubin, B. D. & Benson, J. F. Influence of resource selection on nonbreeding season mortality of mallards. J. Wildl. Manag. 86, 1–12 (2022).
- Fagan, W. F. *et al.* Spatial memory and animal movement. *Ecol. Lett.* 16, 1316–1329. https://doi.org/10.1111/ele.12165 (2013).
 Oliveira-Santos, L. G. R., Forester, J. D., Piovezan, U., Tomas, W. M. & Fernandez, F. A. S. Incorporating animal spatial memory
- in step selection functions. J. Anim. Ecol. 85, 516–524 (2016).
 R. S. Moorgraft, B. P. Orsi, F. & Cagnergi, E. Farguing and anidates of memory based forgoing decisions in a large with
- Ranc, N., Moorcroft, P. R., Ossi, F. & Cagnacci, F. Experimental evidence of memory-based foraging decisions in a large wild mammal. Proc. Nat. Acad. Sci. 118, e2014856118. https://doi.org/10.1073/pnas.2014856118 (2021).
- 112. Bracis, C., Gurarie, E., Rutter, J. & Goodwin, R. A. Remembering the good and the bad: Memory-based mediation of the food-safety trade-off in dynamic landscapes. *Theor. Ecol.* **11**, 305–319. https://doi.org/10.1007/s12080-018-0367-2 (2018).
- Bracis, C. & Wirsing, A. J. Prey foraging behavior after predation introduction is driven by resource knowledge and exploratory tendency. *Front. Ecol. Evol.* 9, 698370. https://doi.org/10.3389/fevo/2021.69830 (2021).
- Poudyal, N. C. & Shrestha, S. Tennessee duck hunter survey 2019–2020. TWRA Wildlife Technical Report 20-4, Nashville, Tennessee, USA (2020).
- Cox, R. R. & Afton, A. D. Use of mini-refuges by female northern pintails wintering in southwestern Louisiana. Wildl. Soc. Bull. 26, 130–137 (1998).
- 116. Rave, D. P. Do mini-refuges supply wintering northern pintails with important diurnal roost sites?. *Wildl. Soc. Bull.* 27, 897–900 (1999).
- Rave, D. P. & Cordes, C. L. Time-activity budgets of northern pintails using nonhunted rice fields in southwest Louisiana. J. Field Ornithol. 64, 211–218 (1993).
- Cox, R. R. & Afton, A. D. Do mini-refuges supply wintering northern pintails with important diurnal roost sites? Response to Rave. Wildl. Soc. Bull. 27, 901–903 (1999).
- 119. Madsen, J. & Fox, A. D. Impacts of hunting disturbance on waterbirds—A review. Wildl. Biol. 1, 193-207 (1995).
- Burger, L. W., Evans, K. O., McConnell, M. D. & Burger, L. M. Private lands conservation: A vision for the future. Wildl. Soc. Bull. 43, 398–407 (2019).
- Nimlos, N. M., Martin, J. A., Palmer, W. E., Sisson, D. C. & Pienaar, E. F. A cost assessment of intensive wild quail management on private lands in the southeastern United States. *Conserv. Sci. Pract.* 5, e12950. https://doi.org/10.1111/csp2.1950 (2023).
 Shumba, T. *et al.* Effectiveness of private land conservation areas in maintaining natural land cover and biodiversity intactness.
- Shumba, T. *et al.* Effectiveness of private land conservation areas in maintaining natural land cover and biodiversity intactness. *Glob. Ecol. Conserv.* 22, e00935 (2020).
 King, S. L., Twedt, D. J. & Wilson, R. R. The role of Wetland Reserve Program conservation efforts in the Mississippi River
- King, S. L., Iwedt, D. J. & Wilson, K. K. The role of Wetland Reserve Program conservation efforts in the Mississippi River Alluvial Valley. Wildl. Soc. Bull. 34, 914–920 (2006).
- 124. Leopold, A. History of the Riley Game Cooperative, 1931–1939. J. Wildl. Manag. 40, 291–302 (1940).
- Adams, K. & Ross, M. QDMA's whitetail report 20171. Quality Deer Management Association (QDMA), 27–29. https://deera ssociation.com/wp-content/uploads/2017/03/WR-2017.pdf. (2017).
- Mitterling, A. M., Rudolph, B. A. & Kramer, D. B. The influence of private land deer management cooperatives on harvest outcomes and hunter satisfaction. Wildl. Soc. Bull. 45, 456–464. https://doi.org/10.1201/9781482295986 (2021).
- Esely, J., Pierce, R. A., Flinn, E. Establishing a wildlife management cooperative. https://extension.missouri.edu/media/wysiw yg/Extensiondata/Pub/pdf/agguides/wildlife/g09490.pdf (2008).
- 128. Rösch, V., Tscharntke, T., Scherber, C. & Batáry, P. Biodiversity conservation across taxa and landscapes requires many small as well as single large habitat fragments. *Oecologia* 179, 209–222 (2015).
- 129. Scharf, A. K., Belant, J. L., Beyer, D. E., Wikelski, M. & Safi, K. Habitat suitability does not capture the essence of animal-defined corridors. *Mov. Ecol.* **6**, 18 (2018).
- Hanski, I. The theories of island biogeography and metapopulation dynamics: Science marches forward but the legacy of good ideas lasts for a long time. In *Theory of Island Biogeography Revisited*. 186–213 http://www.jstor.org/stable/j.ctt7s5m6.12 (Princeton University Press, 2010).
- Han, L. et al. Small patches play a critical role in the connectivity of the Western Tianshan landscape, Xinjiang, China. Ecol. Indic. 144, 109–542. https://doi.org/10.1016/j.ecolind.2022.109542 (2022).
- Burner, R. C., Golas, B. D., Aagaard, K. J., Lonsdorf, E. V. & Thogmartin, W. E. Marginal value analysis reveals shifting importance of migration habitat for waterfowl under a changing climate. *Ecol. Evol.* 13, e10632. https://doi.org/10.1002/ece3.10632 (2023).
- Dayer, A. A., Lutter, S. H., Sesser, K. A., Hickey, C. M. & Gardali, T. Private landowner conservation behavior following participation in voluntary incentive programs: Recommendations to facilitate behavioral persistence. *Conserv. Lett.* 11(2), e12394. https://doi.org/10.1111/conl.12394 (2018).
- 134. Lute, M. L., Gillespie, C. R., Martin, D. R. & Fontaine, J. J. Landowner and practitioner perspectives on private land conservation programs. *Soc. Nat. Resour.* **31**, 218–231 (2018).
- 135. Hanski, I. A practical model of metapopulation dynamics. J. Anim. Ecol. 63, 151-162. https://doi.org/10.2307/5591 (1994).
- 136. Johnson, J. W. Rivers Under Siege: The Troubled Saga of West Tennessee's Wetlands (University of Tennessee Press, 2007).
- U.S. Fish & Wildlife Service. Economic impact of waterfowl hunting in the United States. Addendum to the 2011 National Survey of Fishing, Hunting, and Wildlife-Associated Recreation. Report 2011–1 (2013).
- U.S. Fish & Wildlife Service. Economic impact of waterfowl hunting in the United States. Addendum to the 2011 National Survey of Fishing, Hunting, and Wildlife-Associated Recreation. Report 2011–1 (2015).
- Green, A. W. & Krementz, D. G. Mallard harvest distributions in the Mississippi and Central Flyways. J. Wildl. Manag. 72, 1328–1334. https://doi.org/10.2193/2007-028 (2008).
- Schroeder, S. A., Fulton, D. C., Cornicelli, L., Cordts, S. D. & Lawrence, J. S. Clarifying how hunt-specific experiences affect satisfaction among more avid and less avid waterfowl hunters. *Wildl. Soc. Bull.* 43, 455–467 (2019).
- 141. Stafford, J. D., Pearse, A. T., Hine, C. S., Yetter, A. P. & Horath, M. M. Factors associated with hunter success for ducks on stateowned lands in Illinois, USA. *Wildl. Biol.* **16**, 113–122 (2007).
- 142. Odum, E. P. The Mesocosm. BioScience 34(9), 558–562. https://doi.org/10.2307/1309598 (1984).
- 143. Wu, J. X. *et al.* U.S. National Wildlife Refuge System likely to see regional and seasonal species turnover in bird assemblages under a 2 °C warming scenario. *Ornithol. Appl.* **124**, 016. https://doi.org/10.1093/ornithapp/duac016 (2022).
- 144. Carney, S. M. Species, Age, and Sex Identification of Ducks Using Wing Plumage (U.S. Fish and Wildlife Service, 1992).
- 145. Fair, J., Paul, E. & Jones, J. Guidelines to the Use of Wild Birds in Research 3rd edn. (Ornithological Council, 2010).
- 146. Pebesma, E. Simple features for R: Standardized support for spatial vector data. R J. 10, 439-446 (2018).
- 147. R Core Team. R: A Language and Environment for Statistical Computing http://www.R-project.org/ (R Foundation for Statistical Computing, 2023).
- Alisauskas, R. T., Arnold, T. W., Leafloor, J. O., Otis, D. L. & Sedinger, J. S. Lincoln estimates of mallard (*Anas platyrhynchos*) abundance in North America. *Ecol. Evol.* 4, 132–143. https://doi.org/10.1002/ece3.906 (2014).
- Rohwer, F. C. & Anderson, M. G. Female-based philopatry, monogamy, and the timing of pair formation in migratory waterfowl. In *Current Ornithology* Vol. 5 (ed. Johnston, R. F.) 187–221 (Plenum Press, 1988).
- McDougall, M. B. & Amundson, C. L. Harvest dynamics and annual survival of mallards and gray ducks. J. Wildl. Manag. 81, 449–460. https://doi.org/10.1002/jwmg.21213 (2017).
- Riecke, T. V. *et al.* A hierarchical model for jointly assessing ecological and anthropogenic impacts on animal demography. *J. Anim. Ecol.* 91, 1612–1626. https://doi.org/10.1111/1365-2656.13747 (2022).

- Borg, N. J. et al. Behavioral connectivity among bighorn sheep suggests potential for disease spread. J. Wildl. Manag. 81, 38–45. https://doi.org/10.1002/jwmg.21169 (2017).
- Devineau, O. et al. Increased flexibility for modeling telemetry and nest-survival data using the multistate framework. J. Wildl. Manag. 78, 224–230. https://doi.org/10.1002/jwmg.660 (2014).
- Kéry, M. & Schaub, M. Bayesian Population Analysis Using WinBugs: A Hierarchical Perspective 1st edn. (Academic Press, 2012).
 VonBank, J. A. et al. Winter fidelity, movements, and energy expenditure of Midcontinent Greater White-fronted Geese. Mov.
 - *Ecol.* **9**, 2. https://doi.org/10.1186/s40462-020-00236-4 (2021).
- Filazzola, A. & Cahill, J. F. Jr. Replication in ecology: Identifying challenges and proposing solutions. *Methods Ecol. Evol.* 12, 1780–1792. https://doi.org/10.1111/2041-210X.13657 (2021).
- Kellner, K. jagsUI: A wrapper around 'rjags' to streamline 'JAGS' analyses. R package version 1.5.1. https://CRAN.R-project.org/ package=jagsUI (2019).
- 158. Plummer, M. JAGS Version 4.2.0 (2021).
- Gabry, J., Simpson, D., Vehtari, A., Betancourt, M. & Gelman, A. Visualization in Bayesian workflow. J. R. Stat. Soc. Ser. 182, 389–402. https://doi.org/10.1111/rssa.12378 (2019).
- Zhu, J., Chen, J., Hu, W. & Zhang, B. Big learning with Bayesian methods. Natl. Sci. Rev. 4, 627–651. https://doi.org/10.1093/ nsr/nwx044 (2017).
- 161. Brooks, S. P. & Gelman, A. General methods for monitoring convergence of iterative simulations. J. Comput. Graph. Stat. 7, 434–455 (1998).
- 162. Kruschke, J. Doing Bayesian Data Analysis: A Tutorial with r, JAGS, and stan (Academic Press, 2014).
- 163. McElreath, R. Statistical Rethinking: A Bayesian Course with Examples in r and stan (Chapman, Hall/CRC, 2018).
- Brooks, M. E. et al. glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. R J. 9, 378–400. https://doi.org/10.32614/RJ-2017-066 (2017).
- 165. Burnham, K. P. & Anderson, D. R. Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach 2nd edn. (Springer, 2002).
- Hartig, F. DHARMa: Residual diagnostics for hierarchical (multi-level/mixed) regression models. R package version 0.4.6. https://florianhartig.github.io/DHARMa/ (2022).
- Cade, B. S., Terrell, J. W. & Schroeder, R. L. Estimating effects of limiting factors with regression quantiles. *Ecology* 80, 311–323 (1999).
- Walshe, T., Wintle, B., Fidler, F. & Burgman, M. Use of confidence intervals to demonstrate performance against forest management standards. For. Ecol. Manag. 247, 237–245 (2007).
- 169. Therneau, T. A package for survival analysis in R. R package version 3.4-0 https://CRAN.R-project.org/package=survival (2022)
- 170. Therneau, T. M. & Grambsch, P. M. Modeling Survival Data: Extending the Cox Model (Springer, 2000).
- Benson, J. F., Patterson, B. R. & Mahoney, P. J. A protected area influences genotype-specific survival and the structure of a Canis hybrid zone. *Ecology* 95, 254–264 (2014).
- Dinkins, J. B., Conover, M. R., Kirol, C. P., Beck, J. L. & Frey, S. N. Greater sage-grouse (*Centrocercus urophasianus*) hen survival: Effects of raptors, anthropogenic and landscape features, and hen behavior. *Can. J. Zool.* 92, 319–330. https://doi.org/10.1139/ cjz-2013-0263 (2014).
- 173. Bland, J. M. & Altman, D. G. The logrank test. Stat. Notes 328, 1073. https://doi.org/10.1136/bmj.328.7447.1073 (2004).

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Competing interests

The authors declare no competing interests.

Additional information

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